Supplementary Method 1. Immunohistochemical study.

Immunohistochemical analysis was performed on paraffin tissue sections, which were cut at a thickness of 4 μm. IHC-Tek™ epitope retrieval solution and steamer (IHC world, Woodstock, MD, USA) were used for antigen retrieval of CD42b epitopes. After cooling from the heat, sections were treated with 0.075% glycine in phosphate-buffered saline (PBS) for 10 minutes, and blotto (Tris-buffered saline containing 1% horse serum and 5% skim milk) was added thereafter. Following the blotto incubation, the sections were incubated with the primary antibodies for 2 hours at 37°C (for myeloperoxidase), or overnight at 4°C (for CD42b, fibrinogen, and glycophorin A). The following primary antibodies were used in this study: rabbit monoclonal anti-CD42b (1:100, ab134087; Abcam, Cambridge, UK), rabbit polyclonal anti-Fibrinogen (1:200, ab34269; Abcam), rabbit monoclonal anti-Glycophorin A (1:400, ab129024; Abcam), and mouse monoclonal anti-myeloperoxidase (1:200, MAB3174; R&D systems, Inc., Minneapolis, MN, USA). Sections with blotto lacking a primary antibody were used as a negative control. Next, sections were incubated with secondary antibodies for 30 minutes at 37°C, followed by 0.3% hydrogen peroxide in methyl alcohol for 20 minutes. The sections were then subjected to a 2-minute tap water wash, before being treated with an avidin-biotin-horseradish peroxidase complex (Vector Laboratories Ltd., Peterborough, Cambridgeshire, UK) for 30 minutes to amplify the signals. The signals were visualized by 3,3′-diaminobenzidine solution (PBS containing 0.05% 3,3′-diaminobenzidinetetrahydrochloride hydrate and 0.1% hydrogen peroxide). After counterstaining with hematoxylin, the slides were mounted with Permount mounting medium (Fisher Scientific, Fair lawn, NJ, USA). Sections were viewed using Axio Imager D2 microscope and Axio Vision software (Carl Zeiss MicroImaging GmbH, Jena, Germany).

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